

Poster abstract submission

Approval Status

Not Started

Presenting author

Patrick Moritz

Presenting author's email

patrick.moritz@med.uni-tuebingen.de

Further authors (if any)

Thales Kronenberger
Samuel Wagner

Affiliation(s)

Universitätsklinikum Tübingen, Institut für medizinische Mikrobiologie und Hygiene, Tübingen, Germany
University of Tübingen, Interfaculty Institute of Microbiology and Infection Medicine (IMIT), Tübingen, Germany
German Center for Infection Research (DZIF), Partner-Site Tübingen, Tübingen, Germany

Country

Germany

Type of organization

Academic / research institution

Poster title

Targeting *Salmonella*'s virulence: Inhibition of the T3SS-2 major export apparatus protein SsaV by a synthetic small molecule

Poster abstract

The enteric pathogen *Salmonella enterica* serovar *Typhimurium* utilizes multiple effector proteins to invade and multiply within host epithelial cells, enabling systemic infection. Specialized secretion systems, such as the Type III Secretion System 2 (T3SS-2), encoded on the *Salmonella* Pathogenicity Island 2 (SPI-2), are essential for the translocation of these effector proteins into host cells, supporting intracellular survival, particularly within macrophages. This enables *Salmonella* to evade the host immune response by manipulating vesicle trafficking and maintaining the integrity of *Salmonella*-containing vacuoles (SCVs), thereby promoting bacterial persistence within the host.

The major export apparatus protein SsaV, located at the core of T3SS-2, is crucial for translocation of bacterial effector proteins into host cells. A functional SsaV is essential for *Salmonella* to establish infection and proliferate intracellularly. Therefore, targeting SsaV to block T3SS-2 function could represent an effective strategy to prevent systemic infections.

Through virtual docking, we identified a synthetic small-molecule binder V9. The *in vitro* binding of V9 to SsaV was validated by microscale thermophoresis (MST) using purified nonameric SsaV protein. We then used a split-luciferase host cell invasion assay that showed significantly reduced effector protein injection in V9-treated *Salmonella*. Consistently, Western blot analysis confirmed impaired effector protein secretion upon V9 treatment. Additionally, fluorescence microscopy and colony-forming unit assays in HeLa cells demonstrated reduced intracellular replication of *Salmonella*. To refine our understanding of V9 binding and improve the compounds efficacy, we performed alanine scanning mutagenesis and identified key residues whose substitution significantly impaired V9 binding. These complementary approaches will guide future structure-activity relationship (SAR) studies and hit optimization.

The inhibition mechanism of V9 offers a foundation for developing anti-virulence therapies targeting T3SS-2, with the potential to effectively treat *Salmonella Typhimurium* infections while minimizing the risk of promoting antibiotic resistance.

Research topic

Small molecule therapeutics